



This Journal of Environmental Horticulture article is reproduced with the consent of the Horticultural Research Institute (HRI – www.hriresearch.org), which was established in 1962 as the research and development affiliate of the American Nursery & Landscape Association (ANLA – <http://www.anla.org>).

HRI's Mission:

To direct, fund, promote and communicate horticultural research, which increases the quality and value of ornamental plants, improves the productivity and profitability of the nursery and landscape industry, and protects and enhances the environment.

The use of any trade name in this article does not imply an endorsement of the equipment, product or process named, nor any criticism of any similar products that are not mentioned.

Literature Cited

1. Appleton, B.L. and C.E. Whitcomb. 1983. Using super absorbent to reduce wilting and water requirements of container grown plants. Res. Rept. P-843, Okla. State Univ., Stillwater.
2. Askew, J.C., C.H. Gilliam, H.G. Ponder, and G.J. Keever. 1985. Transplanting leafed-out bare root dogwood liners. HortScience 20:219-221.
3. Baxter, L. and L. Waters, Jr. 1986. Effect of hydrophilic polymer seed coating on the field and laboratory performance of sweet corn (*Zea mays*) and cowpea (*Vigna unguiculata*). J. Amer. Soc. Hort. Sci. 111:31-34.
4. Bearce, B.C. and R.W. McCollum. 1977. A comparison of peatlite and noncomposted hardwood-bark mixes for use in pot and bedding-plant production and the effects of a new hydrogel soil amendment on their performance. Flor. Rev. 161(4169):21-23, 66.
5. Bilderback, T.E. 1987. Moisture extender and wetting agent effects on two drought sensitive nursery crops. HortScience 22:1049.
6. Ferrazza, J. 1974. Grower evaluates soil amendment. Flor. Rev. 155(4019):27, 69-70.
7. Flannery, R.L. and W.J. Busscher. 1982. Use of a synthetic polymer in potting soils to improve water holding capacity. Commun. Soil Sci. Plant Anal. 13:103-111.
8. Gehring, J.M. and A.J. Lewis, III. 1980. Effect of hydrogel on wilting and moisture stress of bedding plants. J. Amer. Soc. Hort. Sci. 105:511-513.
9. Henderson, J.C. and F.T. Davies, Jr. 1987. Effect of a hydrophilic gel on water relations, growth, nutrition of landscape roses. HortScience 22:1114.
10. Henderson, J.C. and D.L. Hensley. 1985. Ammonium and nitrate retention by a hydrophilic gel. HortScience 20:667-668.
11. Henderson, J.C. and D.L. Hensley. 1987. Do hydrophilic gels improve germination and survival? Amer. Nurseryman, Vol. 166 (No. 4): pg. 189-190, 192, 194.
12. Ingram, D.L. and T.H. Yeager. 1987. Effects of irrigation frequency and a water-absorbing polymer amendment on ligustrum growth and moisture retention by a container medium. J. Environ. Hort. 5:19-21.
13. Johnson, M.S. 1984. Effect of soluble salts on water absorption by gel-forming soil conditioners. J. Sci. Food Agric. 35:1063-1066.
14. Rogers, C.S. and R.C. Anderson. 1981. Prairie grass response to strip mine spoil amended with sewage sludge. Bul. Ecol. Soc. Amer. 62:143.
15. Still, S.M. 1976. Growth of 'Sunny Mandalay' chrysanthemums in hardwood-bark-amended media as affected by insolubilized poly(ethylene oxide). HortScience 11:483-484.
16. Taylor, K.C. and R.G. Halfacre. 1986. The effect of hydrophilic polymer on media water retention and nutrient availability to *Ligustrum lucidum*. HortScience 21:1159-1161.
17. Union Carbide Corp. 1973. Agricultural hydrogel, concentrate 50G. Tech. Bul. Creative Agricultural Systems. New York.
18. Wang, Y. 1987. Driving your soil to drink. Greenhouse Mgr. July, pg. 115-121.
19. Whitmore, T.E. 1982. Transplant survival improved. Christmas Trees 10(1):10-11.

Response of Two Florist Azalea Cultivars to Foliar Applications of a Growth Regulator¹

Gary J. Keever² and William J. Foster³

Department of Horticulture
Alabama Agricultural Experiment Station
Auburn University, Auburn, AL 36849

Abstract

Bonzi (paclobutrazol) sprays of 100 and 150 ppm controlled bypass shoot development and increased flower number of 'Alaska' azalea compared to the control, while minimally affecting forcing time and bloom size. Sprays of 150 and 200 ppm suppressed bypass shoot development and increased flower number of 'Prize' azalea compared to the control without affecting bloom size. Paclobutrazol was more effective than B-Nine (daminozide) in suppressing bypass shoot development and enhancing flowering. Forcing time decreased and bloom size increased for paclobutrazol-treated plants of both cultivars compared to daminozide-treated plants.

Index words: growth retardant, bypass shoots

Growth regulators used in this study: Bonzi (paclobutrazol) B-[(4-chlorophenyl) methyl]- α -(1,1-dimethylethyl)-1H-1,2,4-triazole-1-ethanol; B-nine (daminozide) butanedioic acid mono (2,2-dimethylhydrazide).

Introduction

Growth retardants are an accepted component of florist azalea production. In addition to suppressing internode elongation (8), growth retardants promote flower bud initiation

(3), result in multiple flower buds frequently forming on individual shoots (7), and hasten flower development (3). Stuart (8) suggested that growth retardants inhibit the growth of vegetative shoots that develop below the flowers (bypass shoots) and, more recently, growth retardants were shown to suppress bypass shoot development (9).

Daminozide [butanedioic acid mono(2,2-dimethylhydrazide)] and chlormequat chloride [2-chloro-N,N,N-trimethylethanaminium chloride] are the principal growth retardants applied to florist azaleas. Delayed flowering and smaller flower size are undesirable side effects of daminozide (1),

¹Received for publication October 20, 1988; in revised form December 22, 1988.

²Associate Professor of Horticulture.

³Superintendent, Ornamental Horticulture Substation, Mobile, AL 36689. Plants used in this research were provided by Blackwell Nurseries, Semmes, AL 36575.

while delayed flowering and smaller plant size are undesirable effects of chlormequat chloride (5). Paclobutrazol, currently labeled as Bonzi® for use on poinsettia, is an effective retardant on chrysanthemums (6), many species of tropical foliage plants (4) and annual bedding plants (2). This study was conducted to determine the effectiveness of paclobutrazol in controlling bypass shoots of florist azaleas relative to daminozide and to evaluate the chemical's effects on flowering.

Materials and Methods

Uniform 8.9 cm (3.5 in) liners of *Rhododendron* × 'Prize' and 'Alaska' were potted in March 1987, into 1.5 l (6 in) containers of peat:softwood shavings (3:2 by vol) growth medium amended with 3.6 kg/m³ (6 lb/yd³) SREF 19N-1P-8.3K (19-2-10), 3.6 kg/m³ (6 lb/yd³) dolomitic limestone, and 0.4 kg/m³ (0.75 lb/yd³) Micromax. Plants were placed in a double polyethylene greenhouse in a commercial azalea nursery in Semmes, Alabama, and maintained according to common commercial practices. Plants were sheared on July 1 and sprayed the following day with 3627 ppm dikegulac [2,3,4,6-bis-0-(1-methylethylidene)-α-L-xylo-2-hexulofuranosonic acid] to increase lateral branching. Plants were transferred to a shaded double polyethylene greenhouse (30% light exclusion, 20°C (68°F) minimum night temperature) 10 weeks after shearing (September 14) and divided into 8 equal groups. The following treatments were applied on September 15 in a volume of 204 ml/m² (2 qt/100 ft²): single paclobutrazol sprays of 0, 50, 100, 150, 200, 250, and 300 ppm and a daminozide spray of 3,000 ppm repeated 1 week later. Sprays were applied using a hand-held sprayer to uniformly wet foliage and stems. Treatments were applied approximately 0800 hours on clear days. Greenhouse temperature was 20°C (68°F) with 82% relative humidity at time of application. There were 5 replicates of 3 plants each completely randomized within a cultivar.

Plants received a weekly application of 150 ppm N from Peter's 20N-4.3P-16.6K (20-10-10) Peat-lite Special through November 16. Plants were sprayed with a Benlate®/Daconil® mixture [2 ml each/l (1.5 tsp each/gal)] on November

23 and subsequently cooled in the dark at 3.3°C (38°F) for 6 weeks. Plants were removed from the cooler on January 4, 1988, and placed under shade (47% light exclusion) in a glass greenhouse (20°C (68°F) minimum night temperature). Upon removal of plants from the cooler, open blooms were removed from all plants; open bloom number did not vary among treatments (data not shown). After 4 days, the shade cloth was removed and plants were forced in full sun. Time until flowering was determined from the time plants were removed from the cooler until flowers were fully open. At this time, flower number and diameter (3 randomly selected blooms per plant) and bypass shoot number and length (mean length of the 3 longest bypass shoots on each plant) were determined. Rate response to paclobutrazol was determined by regression analysis, and Dunnett's test for least significant differences was used to compare daminozide to each of the other treatments.

Results and Discussion

Bypass shoot number and length of both cultivars decreased as paclobutrazol rate increased (Tables 1 and 2). At or above the 150 ppm rate, essentially no bypass shoots developed on either cultivar. Daminozide was ineffective in controlling bypass shoot number, with daminozide-treated plants forming more bypass shoots than the control plants and plants treated with all rates of paclobutrazol. Daminozide did suppress bypass shoot elongation of 'Alaska' but not of 'Prize'.

Increasing rates of paclobutrazol from 0 to 300 ppm resulted in linear increases in days to flowering, from 50.3 days to 55.5 days for 'Alaska' plants and from 42.9 days to 48.5 days for 'Prize' plants. Daminozide-treated plants of both cultivars required longer to reach full bloom than untreated plants and plants treated with all rates of paclobutrazol (Fig. 1). There was a quadratic effect of increasing paclobutrazol rate on flower number per plant. Flower number of 'Alaska' increased from 194.7 for the control to 225.1 for plants treated with 100 ppm paclobutrazol but decreased to 201.4 for plants receiving a 300 ppm paclobutrazol spray. Flower number of 'Prize' increased from 138.2 for the con-

Table 1. Paclobutrazol effects on bypass shoot and flower development of *Rhododendron* × 'Alaska'.

Growth retardant	Treatment Concentration (ppm)	Bypass shoots		Days to open flower [†]	Flowers	
		Number	Length [‡] (cm)		Number	Diameter [§] (cm)
Paclobutrazol	0	7.5* ^w	7.9*	50.3*	194.7	6.7*
	50	3.0*	4.3	52.4*	217.7*	6.4*
	100	0.9*	4.5	52.0*	225.1*	6.6*
	150	0.0*	0.0*	53.0*	220.1*	6.5*
	200	0.0*	0.0*	55.0*	216.5*	6.2*
	250	0.0*	0.0*	54.2*	199.1	6.3*
	300	0.0*	0.0*	55.5*	201.5	6.2*
Significance of rate ^v		c	c	l	q	l
Daminozide	3000	9.2	4.3	67.1	177.7	5.5

[‡]Mean length of 3 longest bypass shoots on each plant.

[†]Days to full bloom beginning when plants moved from cooler to greenhouse.

[§]Mean of 3 randomly selected blooms per plant.

^wDunnett's test for least significant differences; means followed by an asterisk differ significantly from the mean of the daminozide treatment at the 5% level.

^vControl included in regression analyses; l = linear, q = quadratic, c = cubic.

Table 2. Paclobutrazol effects on bypass shoot and flower development of *Rhododendron* × 'Prize'.

Growth retardant	Treatment Concentration (ppm)	Bypass shoots			Flowers	
		Number	Length ^z (cm)	Days to open flower ^y	Number	Diameter ^x (cm)
Paclobutrazol	0	5.7* ^w	6.4	42.9*	138.2	7.2*
	50	3.0*	3.9	44.6*	139.1	7.1*
	100	1.3*	3.6	44.6*	144.2	7.1*
	150	0.9*	0.0*	44.3*	147.1*	7.1*
	200	0.0*	0.0*	46.6*	157.0*	7.1*
	250	0.1*	0.0*	46.7*	143.6	7.1*
	300	0.0*	0.0*	48.5*	139.2	7.2*
Significance of rate ^v		q	c	l	q	ns
Daminozide	3000	7.5	5.1	56.5	129.1	6.2

^zMean length of 3 longest bypass shoots on each plant.

^yDays to full bloom beginning when plants moved from cooler to greenhouse.

^xMean of 3 randomly selected blooms per plant.

^wDunnett's test for least significant differences; means followed by an asterisk differ significantly from the mean of the daminozide treatment at the 5% level.

^vControl included in regression analyses; l = linear, q = quadratic, c = cubic.

trol to 157.0 for plants treated with 200 ppm paclobutrazol but dropped to 139.2 at the highest rate of paclobutrazol. Flower number of daminozide-treated 'Alaska' plants was similar to untreated plants and to plants sprayed with 250 and 300 ppm paclobutrazol, but less than that of plants receiving 50, 100, 150, and 200 ppm sprays. Flower number of daminozide-treated 'Prize' plants was less than that of plants treated with 150 and 200 ppm paclobutrazol; flower number of control plants and plants receiving other rates of paclobutrazol was similar to flower number of daminozide-treated plants.

The effect of paclobutrazol on flower diameter varied with cultivar. Flower diameter of 'Alaska' decreased linearly from 6.7 cm (2.6 in) for the untreated control to 6.2 cm (2.4 in) for plants receiving the highest rate of paclobutrazol; flower diameter of 'Prize' was not affected by paclobutrazol rate. Daminozide reduced flower size of 'Alaska' and 'Prize' to less than that of the control and paclobutrazol-treated plants.

Paclobutrazol rates of 100 and 150 ppm applied 5½ weeks before cooling effectively controlled bypass shoot development and increased flower number of 'Alaska' compared to an untreated control, while minimally influencing days-to-flower and flower diameter. Rates of 150 and 200 ppm were most effective in controlling bypass shoots and increasing flower number of 'Prize', while not reducing flower diameter. Days-to-flower was greater at the 200 ppm rate compared to 150 ppm (46.6 vs 44.3). Daminozide was less effective than paclobutrazol in controlling bypass shoot development and enhancing flower number. Daminozide also delayed flowering and reduced flower size relative to paclobutrazol and the untreated control. The delay in flowering and smaller flower size of daminozide-treated plants has been reported previously (1), while the control of bypass shoots with paclobutrazol concurs with recent results by Whealy *et al.* (9).

Significance to the Nursery Industry

Bonzi (paclobutrazol) has the potential of becoming the standard growth retardant for florist azaleas due to superior bypass shoot control and enhanced flowering compared to daminozide. Paclobutrazol-treated plants flowered sooner and had larger blooms than B-Nine (daminozide) treated plants. Optimum Bonzi (paclobutrazol) rates ranged from 150 to 200 ppm for 'Alaska' and 'Prize', but may differ for other cultivars.

(Ed. note: This paper reports the results of research only, and does not imply registration of a pesticide under amended FIFRA. Before using any of the products mentioned in this research paper, be certain of their registration by appropriate state and/or federal authorities.)

Literature Cited

1. Anon. 1969. Yoder azaleas 1970. Growers Circle News 72:3.
2. Barrett, J.E. and T.A. Nell. 1986. Evaluation of XE-1019 and paclobutrazol for height control of flowering annuals. Proc. Plant Growth Reg. Soc. Amer. 13:62-64.



Fig. 1. *Rhododendron* × 'Alaska' treated with 0, 50, 100, and 150 ppm Bonzi (paclobutrazol) [labeled PP 333] and 3000 ppm daminozide (B-nine). Note the bypass shoots on the check plant and the delayed flowering of the B-nine treated plant.

3. Criley, R.A. 1969. Effects of short photoperiods, cycocel, and gibberellic acid upon flower bud initiation and development in azalea 'Hexe'. J. Amer. Soc. Hort. Sci. 94:392-396.

4. Davis, T.P., K. Emino, W. Shurtleff, and N. Sankhla. 1985. The promise of paclobutrazol. Interior Lands. Indus. 2(11):36-41.

5. Larson, R.A. 1975. Continuous production of flowering azaleas. p. 72-77. In: A.M. Kofranek and R.A. Larson (eds.). Growing Azaleas Commercially, Davis, CA. Univ. Calif. Div. of Agr. Sc., Pub. #4058.

6. McDaniel, G.L. 1983. Growth retardation activity of paclobutrazol on chrysanthemum. HortScience 18:199-200.

7. Shanks, J.B. and C.B. Link. 1968. Some factors affecting growth and flower initiation of greenhouse azaleas. Proc. Amer. Soc. Hort. Sci. 92:603-614.

8. Stuart, N.W. 1961. Initiation of flower buds in rhododendron after application of growth retardants. Science 134:50-52.

9. Whealy, C.A., T.A. Nell, and J.E. Barrett. 1988. Plant growth regulator reduction of bypass shoot development in azalea. HortScience 23:166-167.

Evaluation of Nursery Container Designs for Minimization or Prevention of Root Circling¹

Bonnie L. Appleton²

Department of Horticulture
Virginia Polytechnic Institute & State University
Hampton Roads Agricultural Experiment Station
Virginia Beach, VA 23455

Abstract

Six different container designs were evaluated for their effectiveness in minimizing or preventing the development of circling roots around the sides and/or bottoms of containers. While both shrub and tree species were tested, the roots of the shrubs were generally small and fibrous enough that selection of a container to minimize or prevent circling roots was not an important consideration. For the tree species, the greatest amount of circling reduction was achieved with the soft polybags and the rigid stepped-pyramid containers. Because considerable difference exists in the cost of the newly-designed containers, both cost and root-modifying effectiveness should be considered if root modification is deemed important.

Index words: root modification, girdling roots, container-grown trees, poly bags, stepped-pyramid pot, low profile container, ribbed container

Species used in this study: goldenraintree (*Koelreuteria paniculata*); black willow (*Salix nigra*); white pine (*Pinus strobus*); American boxwood (*Buxus microphylla*); azalea (*Rhododendron obtusum* 'Hershey's Red'); honeysuckle privet (*Lonicera pileata*).

Introduction

Design and appearance are factors considered when a grower selects containers for nursery stock production, although the three major selection criteria are generally ease-of-handling, rugged construction and price (5, 10). In addition, features receiving considerable attention lately include color (4), pot lip shape (15), and design for improved winter protection (11).

All aspects of container design influence plant development and growth (and possibly sales). The number of different containers introduced onto the market in recent years has raised the questions of whether standardization of containers is needed, and whether standardization would be beneficial to both wholesale growers and retailers (1, 2, 3, 7). If an effort to standardize is started it could influence the willingness of growers to purchase containers with special design features unless these features are shown to be beneficial to the production of high quality nursery stock.

An additional reason container design, and more specifically side wall configuration, is considered by growers is because of the circling and potentially girdling roots that may develop on certain plants when they are grown in conventional round, smooth, straight-walled rigid containers. Circling roots formed during production have the potential, especially on trees, to enlarge to the point that they may shorten a plant's life span by girdling its stem (6). In addition, circling roots may fail to adequately anchor plants, and may restrict water and nutrient absorption (13).

Research has demonstrated that certain modifications of the container side wall will minimize or prevent circling roots (5, 13, 14). The purpose of this research was to compare the ability of several new container designs to minimize or prevent root circling.

Materials and Methods

Rooted liners of American boxwood (*Buxus microphylla*), black willow (*Salix nigra*), 'Hershey Red' azalea (*Rhododendron obtusum* 'Hershey Red'), and privet honeysuckle (*Lonicera pileata*), and seedlings of goldenraintree (*Koelreuteria paniculata*) and Eastern white pine (*Pinus strobus*),

¹Received for publication September 27, 1988; in revised form December 28, 1988.

²Assistant Professor of Horticulture.